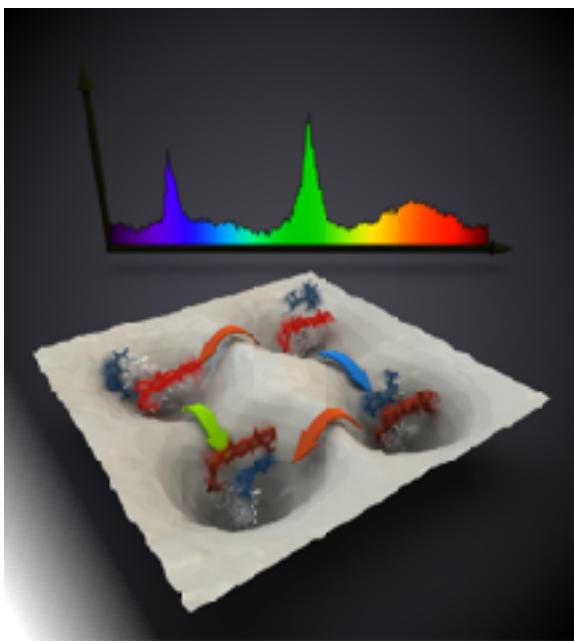


# Accuracy of Markov State Models (MSMs) and Hidden Markov Models (HMMs)



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# Discretization

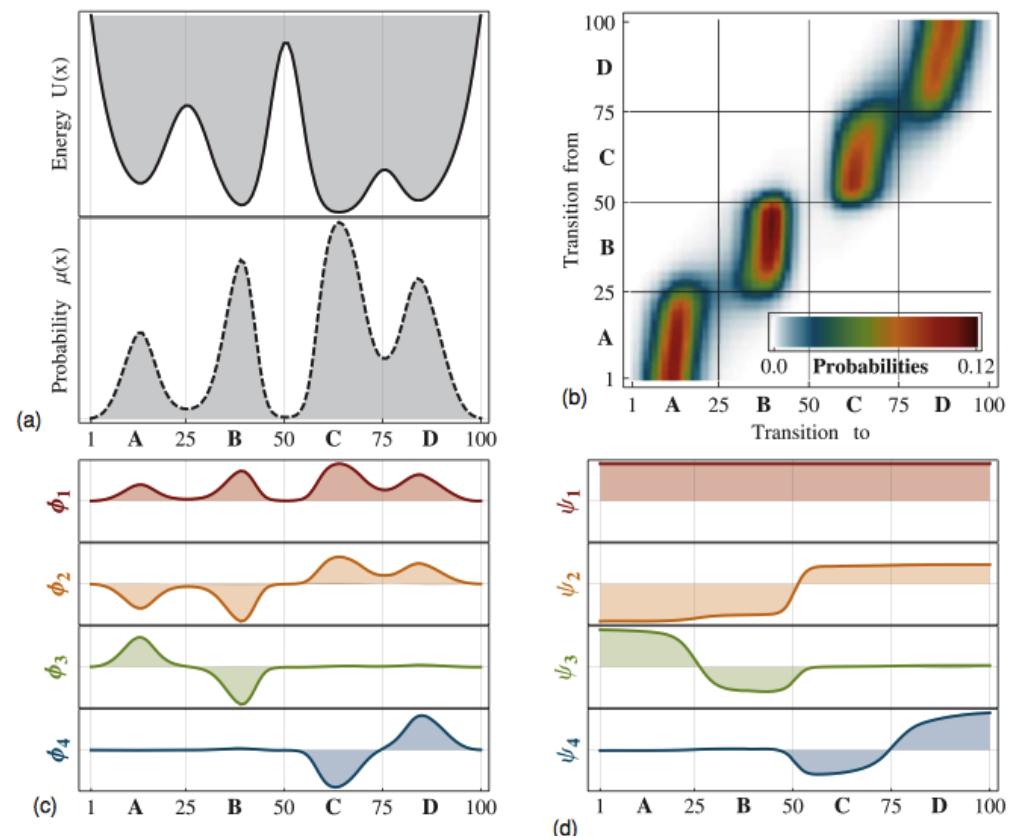
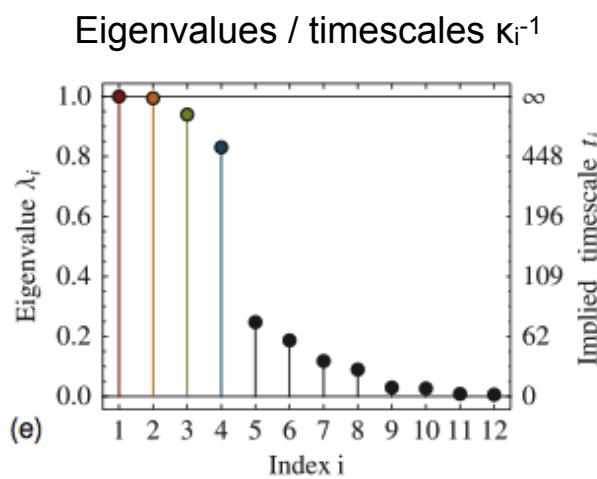
Backward propagator

$$\rho_\tau = \mathcal{T}(\tau)\rho_0$$

Spectral decomposition

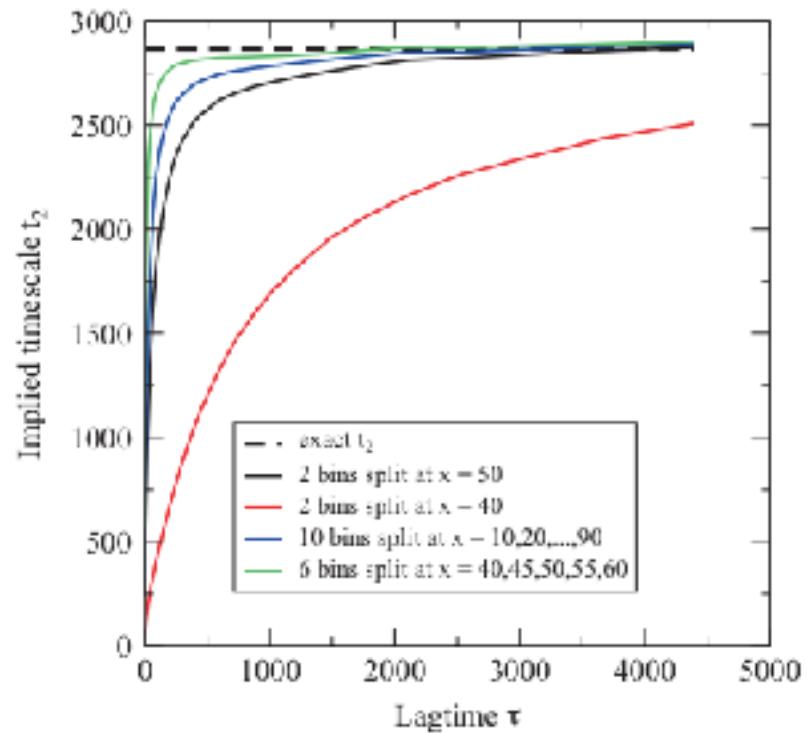
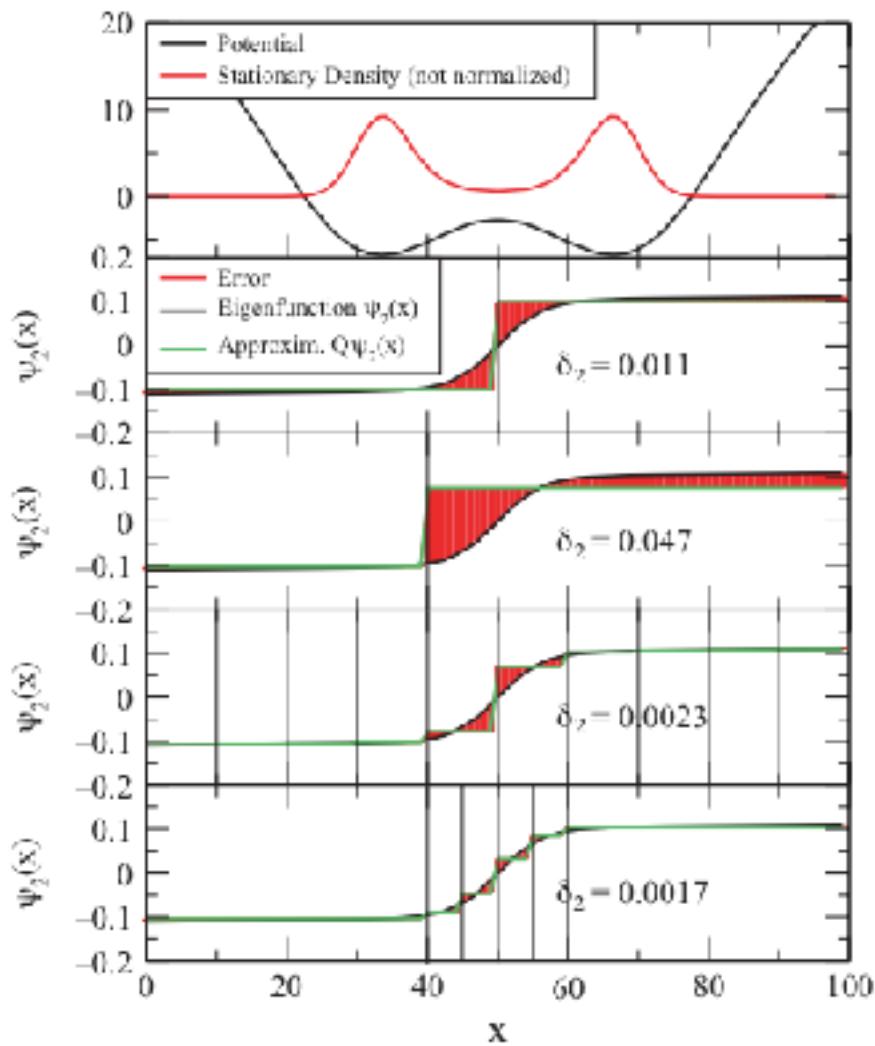
$$\rho_\tau = \sum_{i=1}^{\infty} e^{-\tau \kappa_i} \langle \psi_i | \rho_0 \rangle \psi_i$$

Processes:



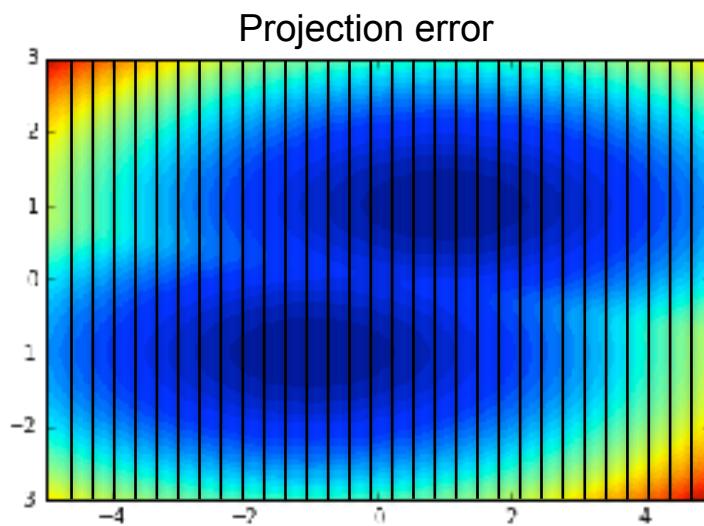
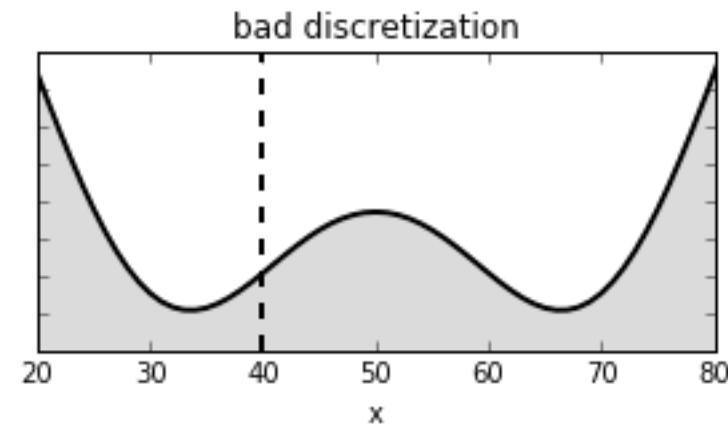
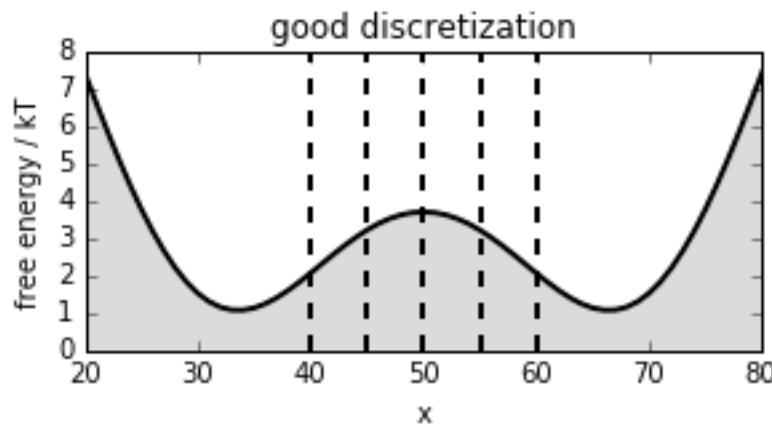
Prinz et al, JCP 134, 174105 (2011)

# Discretization



Prinz et al, JCP 134, 174105 (2011)

# Two flavors of discretization errors



# Projected and hidden Markov models for calculating kinetics and metastable states of complex molecules

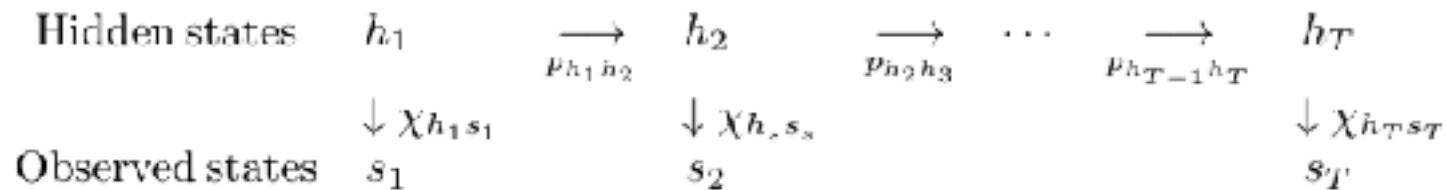
Frank Noé, Hao Wu, Jan-Hendrik Prinz, and Nuria Plattner

Citation: *J. Chem. Phys.* **139**, 184114 (2013); doi: 10.1063/1.4828816

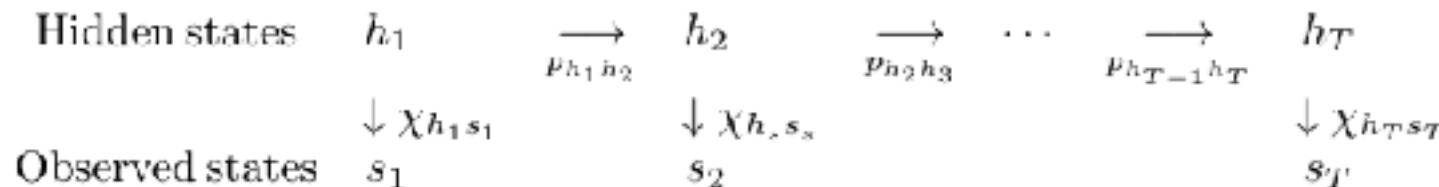
Markov state models (MSMs) have been successful in computing metastable states, slow relaxation timescales and associated structural changes, and stationary or kinetic experimental observables of complex molecules from large amounts of molecular dynamics simulation data. However, MSMs approximate the true dynamics by assuming a Markov chain on a clusters discretization of the state space. This approximation is difficult to make for high-dimensional biomolecular systems, and the quality and reproducibility of MSMs has, therefore, been limited. Here, we discard the assumption that dynamics are Markovian on the discrete clusters. Instead, we only assume that the full phase-space molecular dynamics is Markovian, and a projection of this full dynamics is observed on the discrete states, leading to the concept of Projected Markov Models (PMMs). Robust estimation methods for PMMs are not yet available, but we derive a practically feasible approximation via Hidden Markov Models (HMMs). It is shown how various molecular observables of interest that are often computed from MSMs can be computed from HMMs/PMMs. The new framework is applicable to both, simulation and single-molecule experimental data. We demonstrate its versatility by applications to educative model systems, a 1 ms Anton MD simulation of the bovine pancreatic trypsin inhibitor protein, and an optical tweezer force probe trajectory of an RNA hairpin.

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# Hidden Markov Model



# Hidden Markov Model



Maximum-likelihood Estimation: We define the forward and backward variables

$$\begin{aligned}\alpha_{t,i} &= \mathbb{P}(s_1, \dots, s_t, h_t = i \mid \pi) \\ \beta_{t,i} &= \mathbb{P}(s_{t+1}, \dots, s_T \mid h_t = i)\end{aligned}$$

and the observation matrices

$$\mathbf{O}_t = \text{diag}(\chi_{1,s_t}, \dots, \chi_{m,s_t})$$

The maximum-likelihood estimator is obtained by iterating:

1. **Expectation step:** Estimate the forward-backward variables

$$\boldsymbol{\alpha}_{1:T}, \boldsymbol{\beta}_{1:T} = \arg \max \mathbb{P}(s_{1:T} \mid \boldsymbol{\alpha}_{1:T}, \boldsymbol{\beta}_{1:T}, \tilde{\mathbf{P}}, \boldsymbol{\chi})$$

and compute likelihood

$$L = \log \mathbb{P}(s_{1:T} \mid \tilde{\mathbf{P}}, \boldsymbol{\chi})$$

2. **Maximization step:** Estimate the parameters

$$\tilde{\mathbf{P}}, \boldsymbol{\chi} = \arg \max \mathbb{P}(s_{1:T} \mid \tilde{\mathbf{P}}, \boldsymbol{\chi}, \boldsymbol{\alpha}_{1:T}, \boldsymbol{\beta}_{1:T})$$

## Expectation step:

We estimate the **forward variables**

$$\begin{aligned}\alpha_1 &= \pi \mathbf{O}_1 \\ \alpha_t &= \alpha_{t-1} \tilde{\mathbf{P}} \mathbf{O}_t\end{aligned}$$

and the **backward variables**:

$$\begin{aligned}\beta_T &= 1 \\ \beta_t &= \tilde{\mathbf{P}} \mathbf{O}_t \beta_{t+1}\end{aligned}$$

Maximization step:

Hidden transition count  $c_{t,ij} = \mathbb{P}(h_t = i, h_{t+1} = j | \tilde{\pi}, \tilde{\mathbf{P}}, \chi)$

$$c_{t,ij} = \frac{\alpha_{t,i} p_{ij} \chi_{j,s_{t+1}} \beta_{t+1,j}}{\sum_k \alpha_{t,k}}$$

Hidden state probabilities:

$$\gamma_{t,i} = \frac{\alpha_{t,i} \beta_{t,i}}{\sum_j \alpha_{t,j} \beta_{t,j}}$$

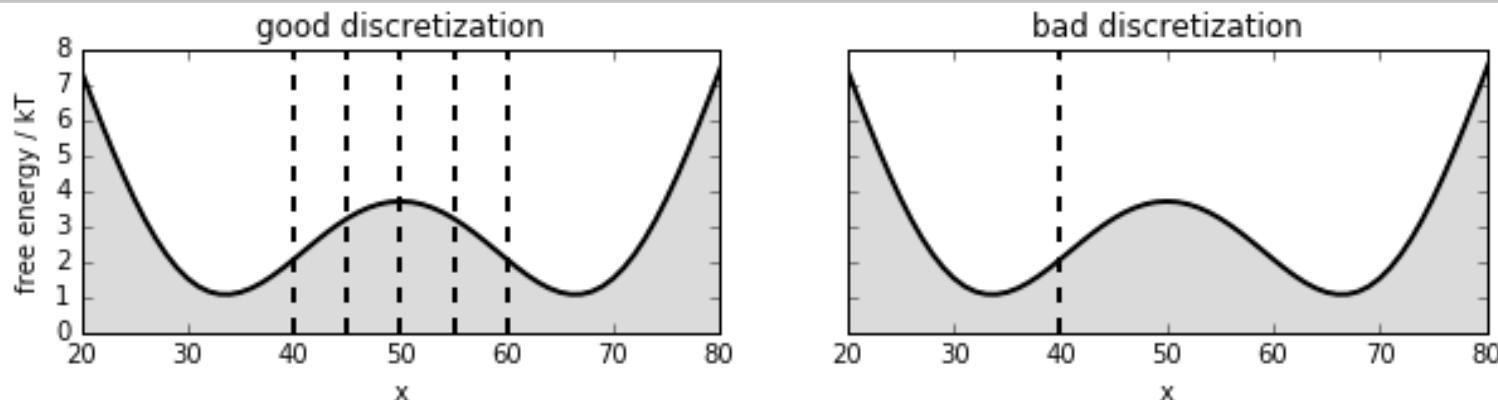
Hidden transition matrix:

$$\tilde{p}_{ij} = \frac{\sum_t c_{t,ij}}{\sum_t \gamma_{t,i}}$$

Hidden initial distribution:

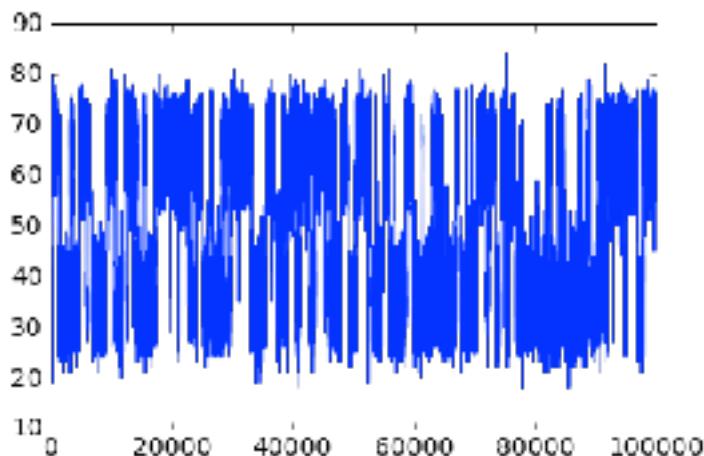
$$\tilde{\pi}_i = \gamma_{1,i}$$

# Case 1: coarse discretization

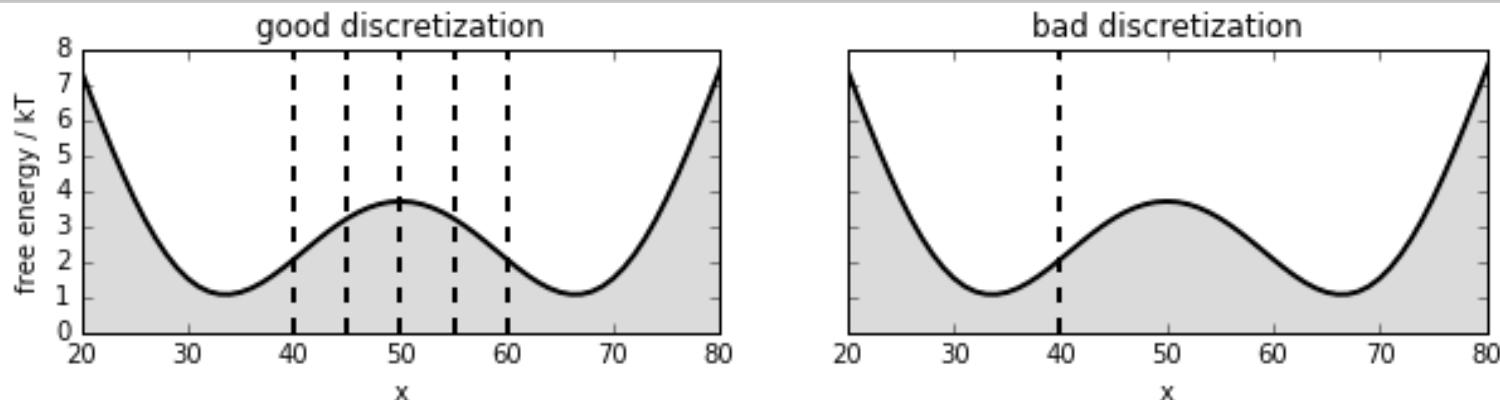


```
# load double well data
import pyemma.datasets
double_well_data = pyemma.datasets.load_2well_discrete()

plot(double_well_data.dtraj_T100K_dt10)
```

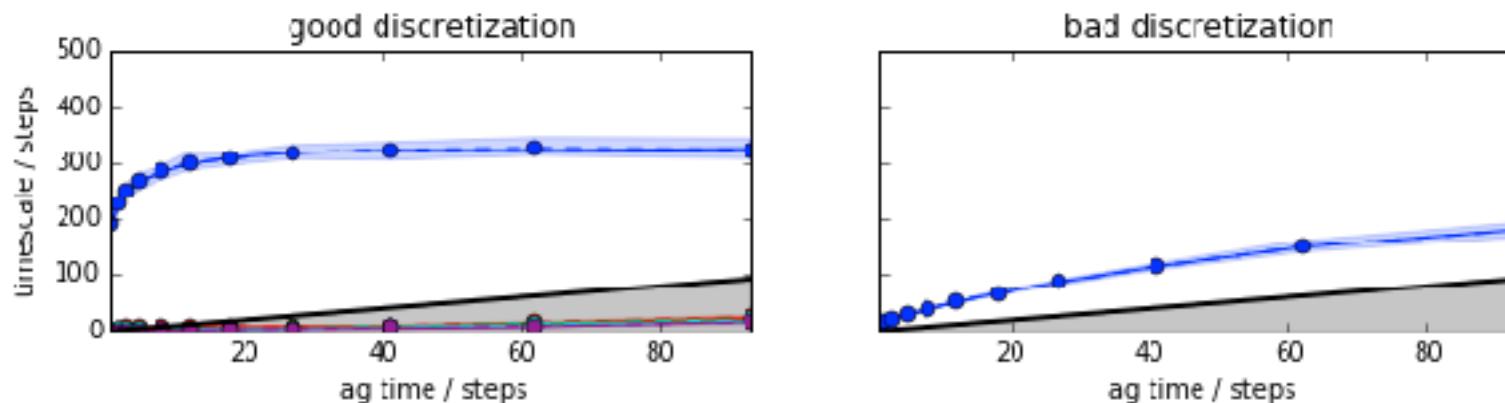


# Case 1: coarse discretization

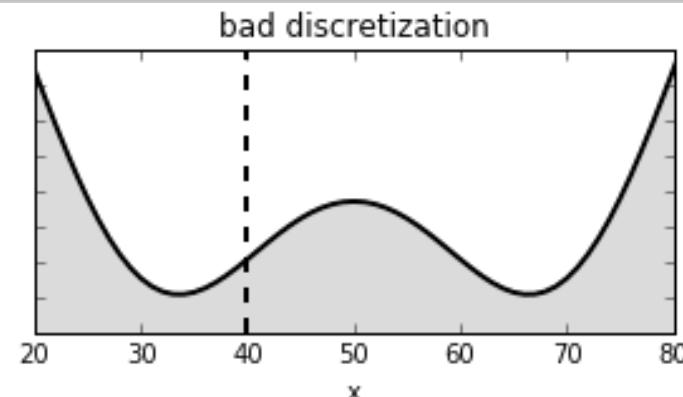
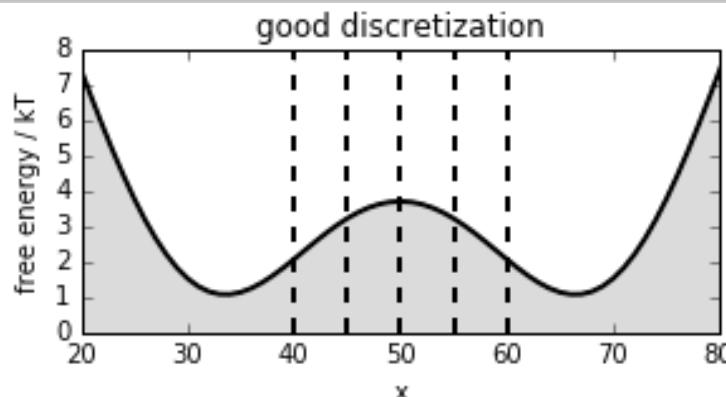


## MSM: Implied timescales

```
its_good_bmsm = msm.timescales_msm([double_well_data.dtraj_T100K_dt10_n6good], lags = 100, errors='bayes')
its_bad_bmsm = msm.timescales_msm([double_well_data.dtraj_T100K_dt10_n2bad], lags = 100, errors='bayes')
```

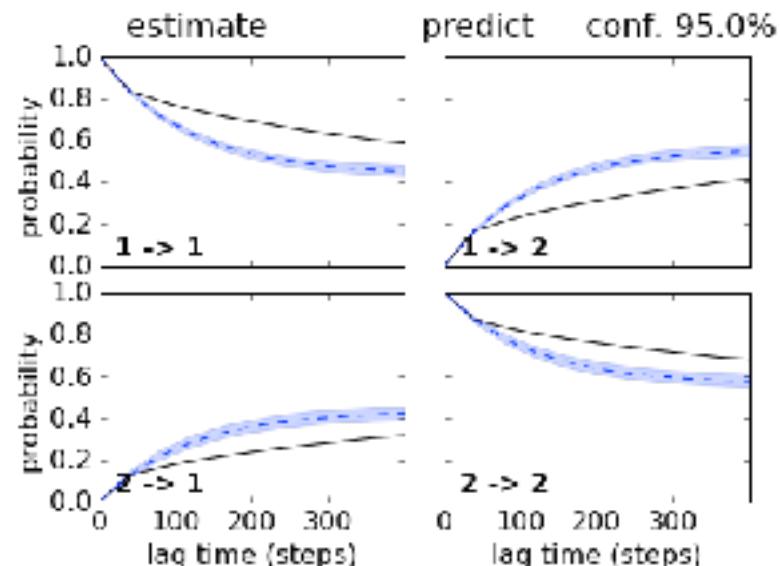
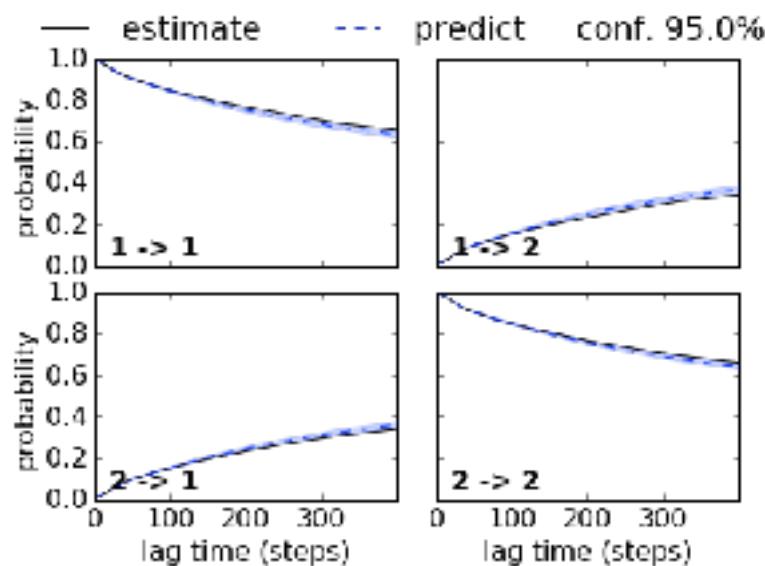


# Case 1: coarse discretization

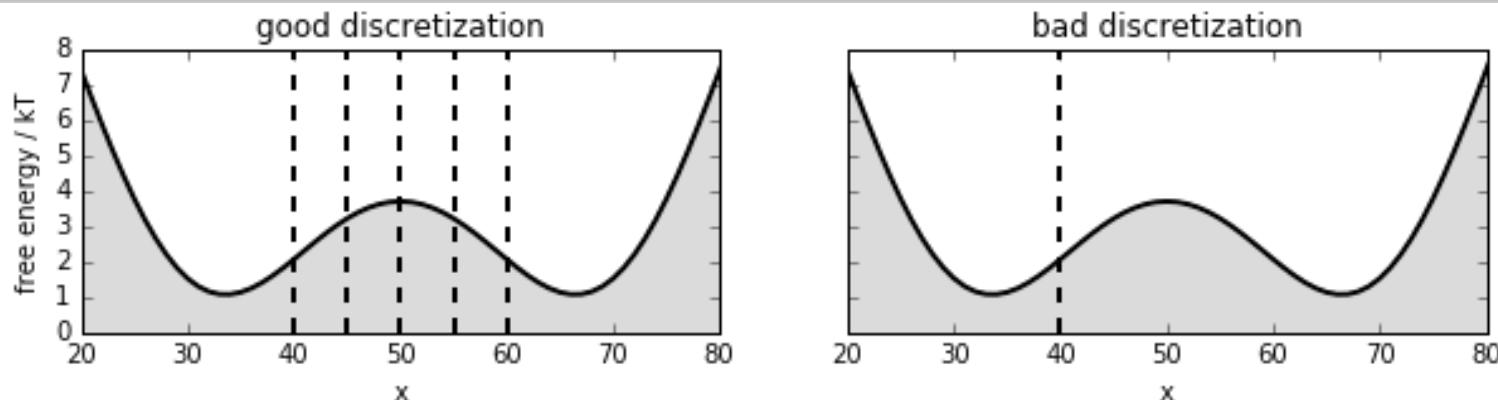


## MSM: Chapman-Kolmogorov Test

```
BMSM_good = msm.bayesian_markov_model([double_well_data.dtraj_T100K_dt10_n6good], 40)
ck_good_bmsm = BMSM_good.cktest(2, mlags=11)
BMSM_bad = msm.bayesian_markov_model([double_well_data.dtraj_T100K_dt10_n2bad], 40)
ck_bad_bmsm = BMSM_bad.cktest(2, mlags=11)
```

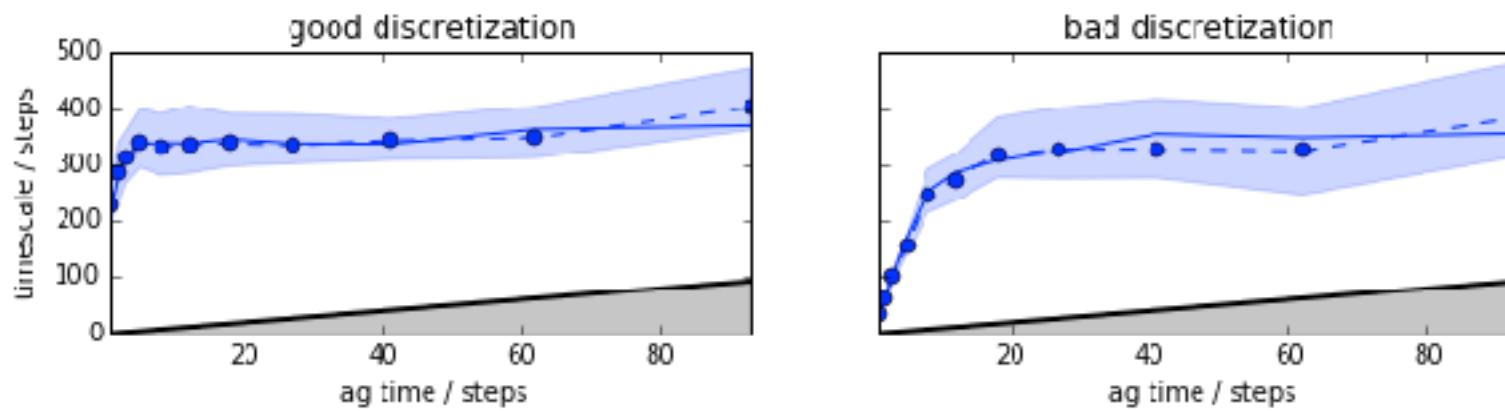


# Case 1: coarse discretization

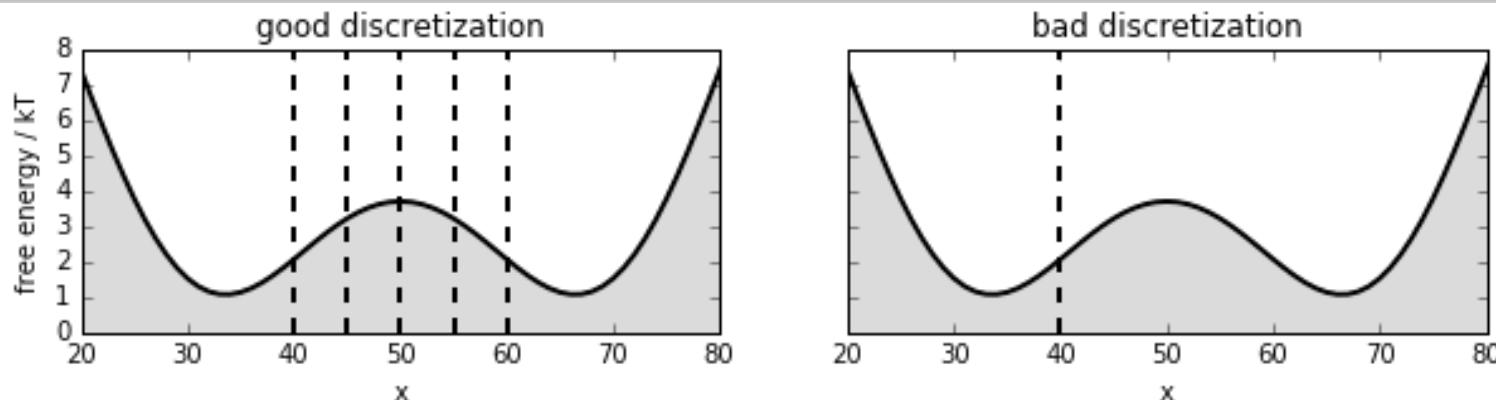


## HMM: Implied timescales

```
its_good_bhmm = mmm.timescales_hmm([double_well_data.dtraj_T100K_dt10_n6good], 2, lags = 100, errors='bayes')
its_bad_bhmm = mmm.timescales_hmm([double_well_data.dtraj_T100K_dt10_n2bad], 2, lags = 100, errors='bayes')
```

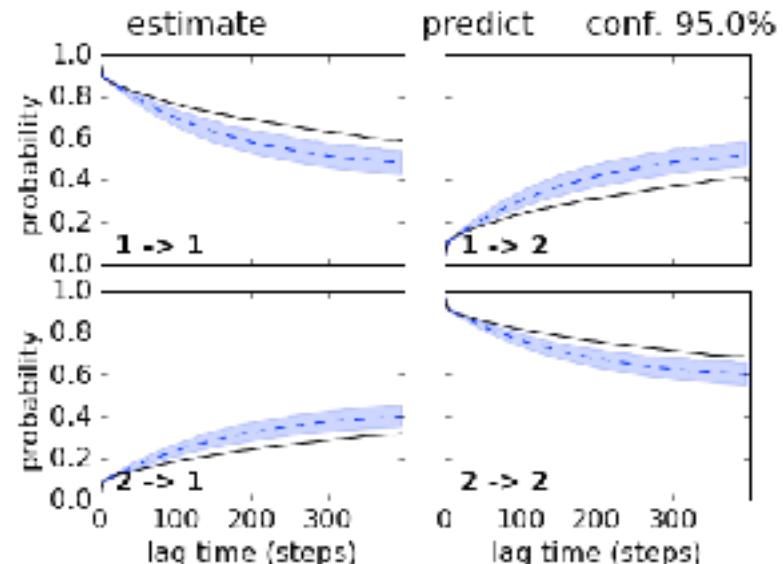
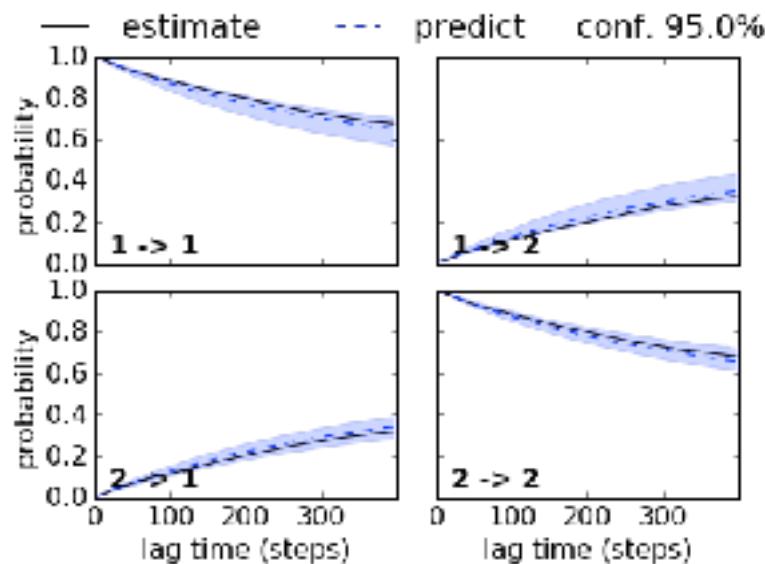


# Case 1: coarse discretization



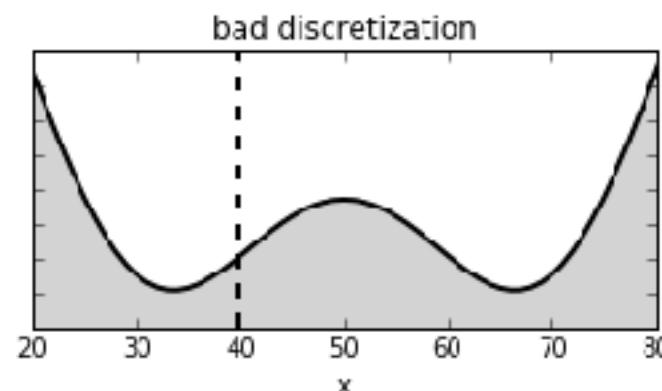
## HMM: Chapman-Kolmogorov Test

```
BHMM_good = msm.bayesian_hidden_markov_model([double_well_data.dtraj_T100K_dt10_n6good], 2, 5)
ck_good_bhmm = BHMM_good.cktest(mlags=80)
BHMM_bad = msm.bayesian_hidden_markov_model([double_well_data.dtraj_T100K_dt10_n2bad], 2, 5)
ck_bad_bhmm = BHMM_bad.cktest(mlags=80)
```

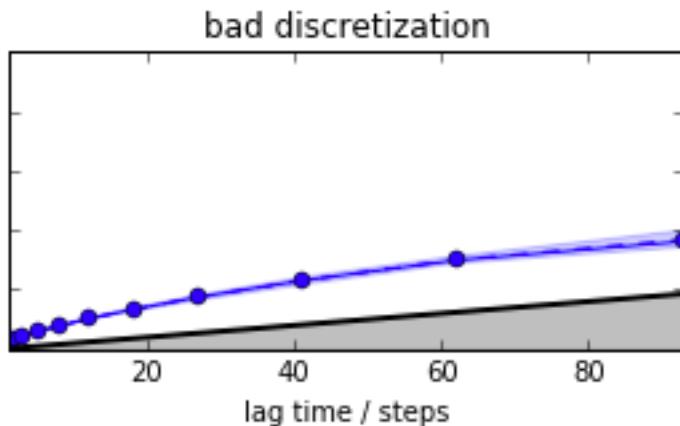


# Case 1: coarse discretization

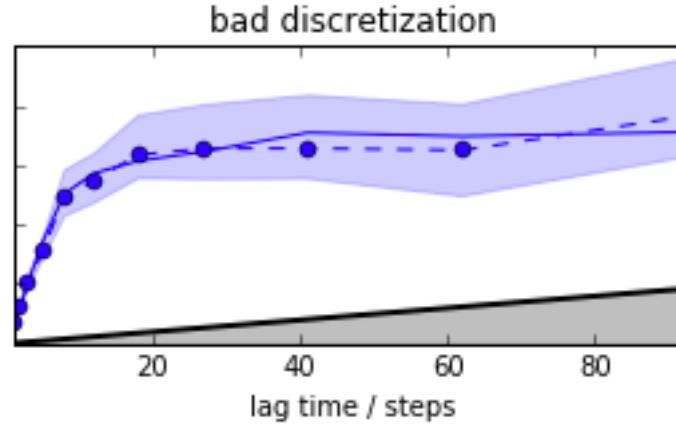
Why does the HMM work better than the MSM?



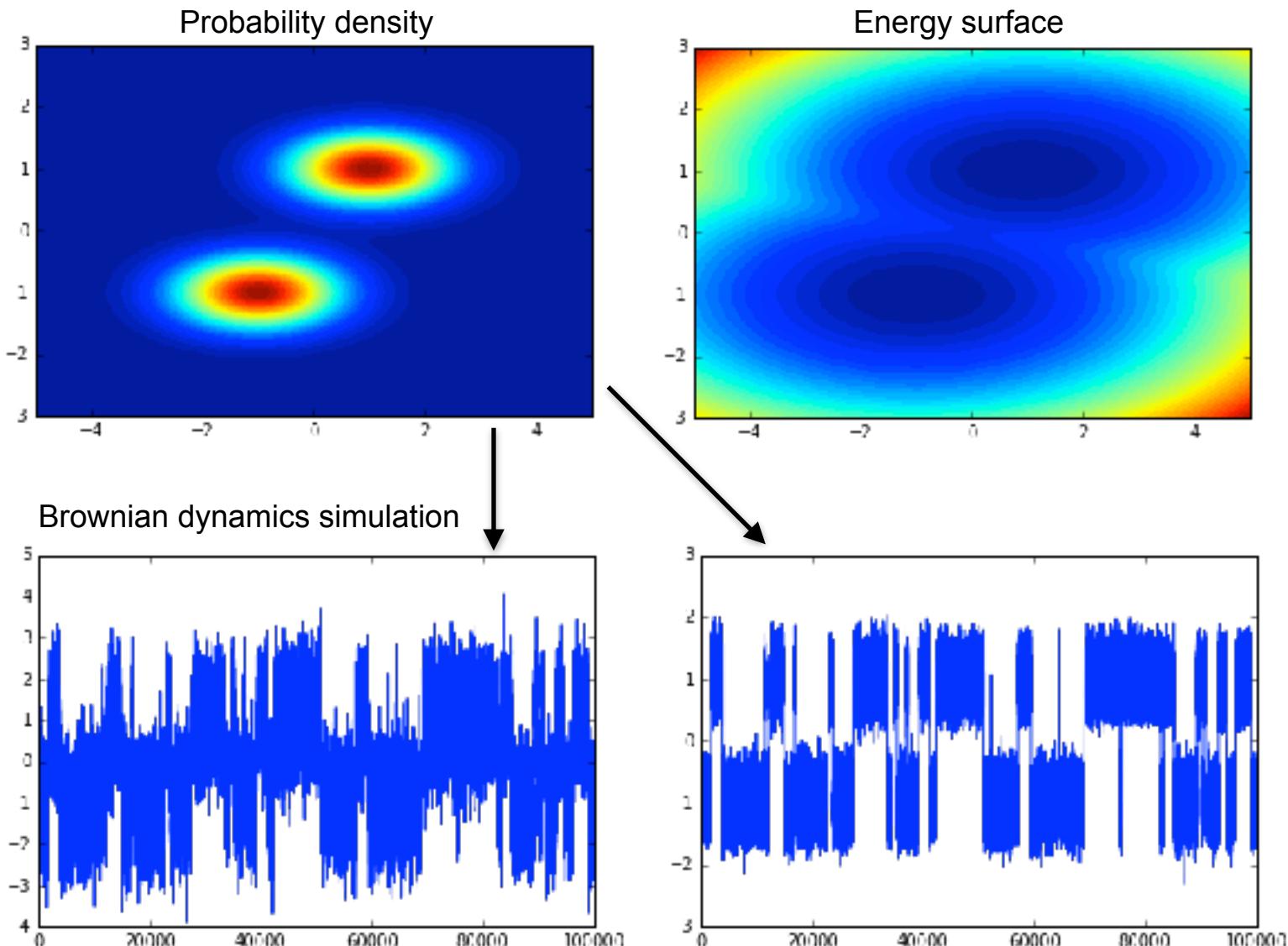
MSM at lag 40:



HMM at lag 40:



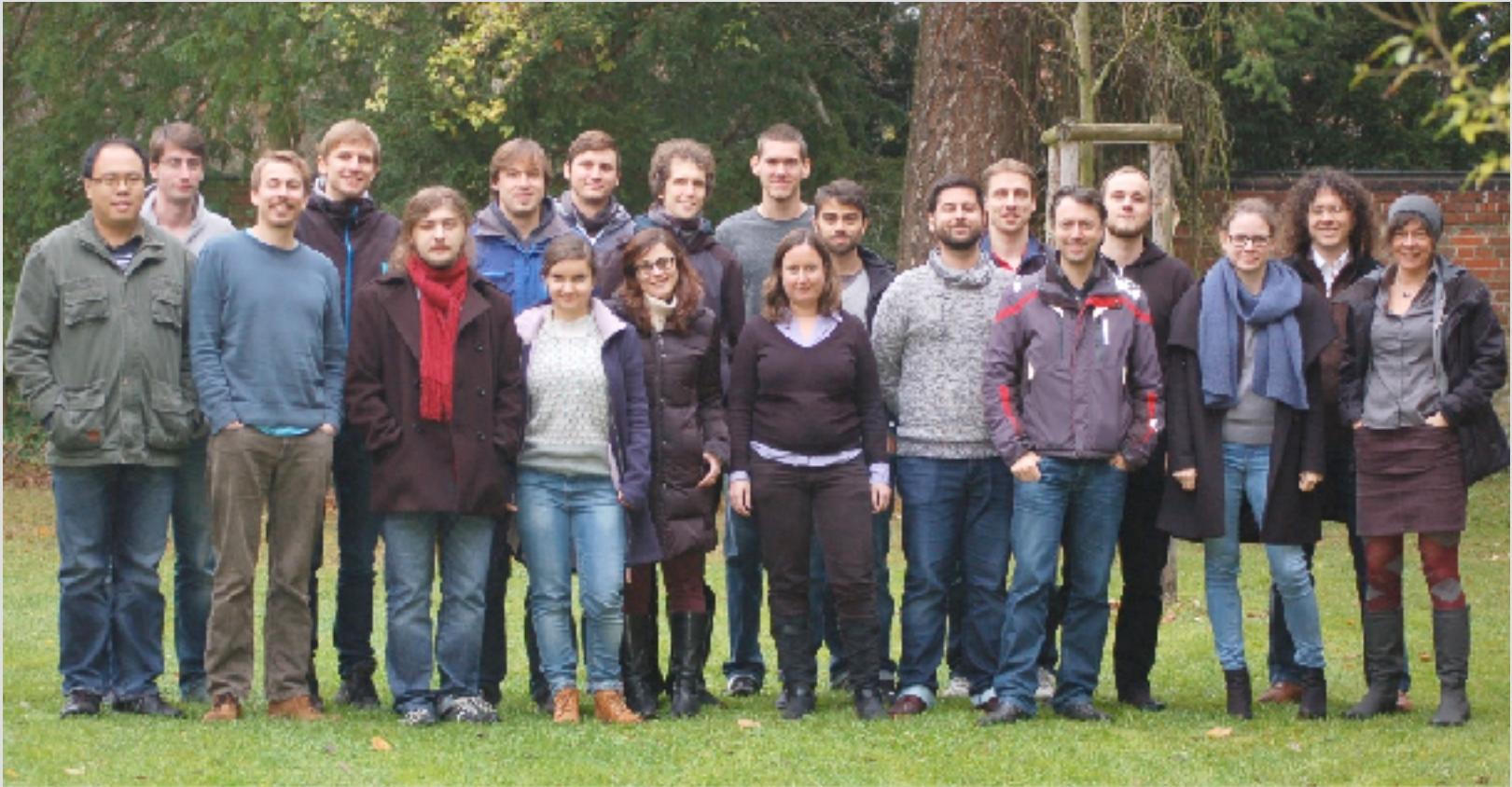
## Case 2: bad projection



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